



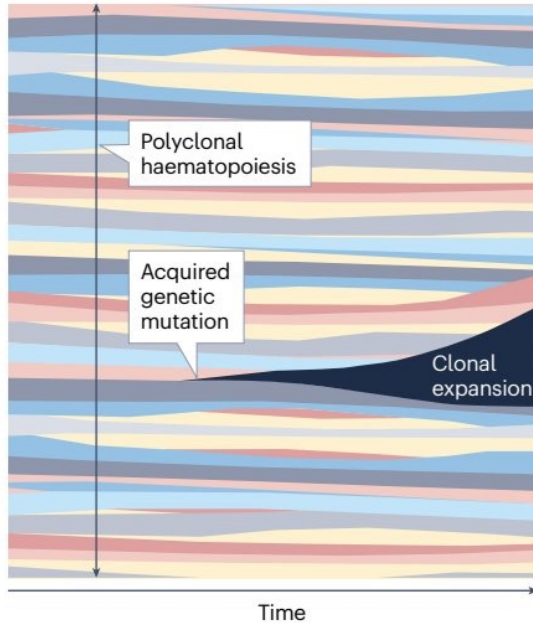
# Clonal hematopoiesis in solid tumor patients: implications for secondary malignancies and diagnostics

***Ahmet Zehir, PhD***

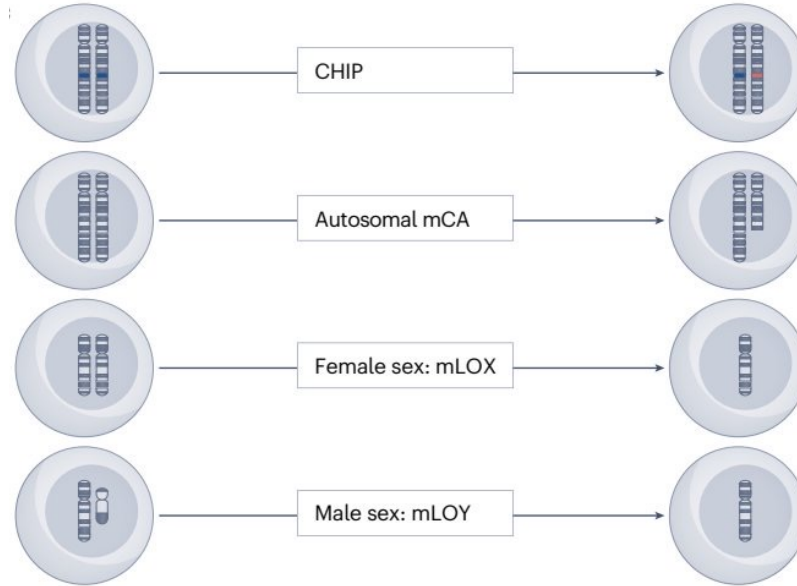
Executive Director, Precision Medicine  
Precision Medicine & Biosamples  
AstraZeneca

# What is clonal hematopoiesis?

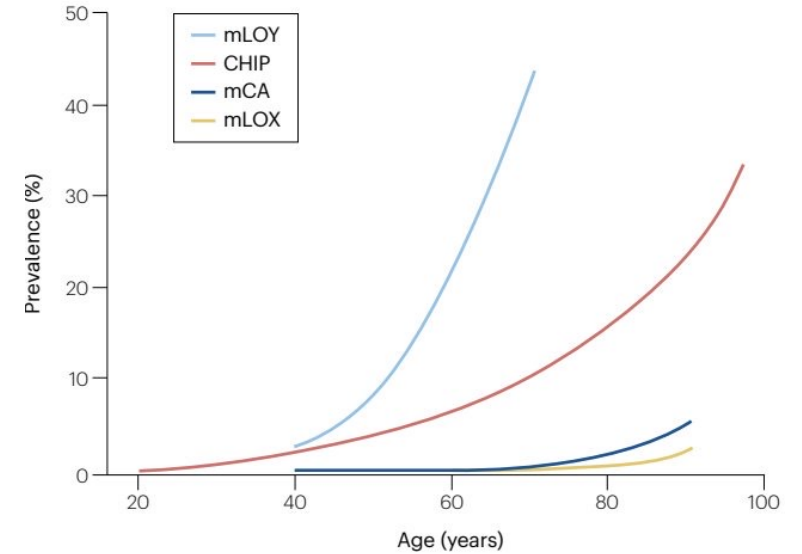
Clonal expansion of hematopoietic (stem) cells with genomic changes that increase cellular fitness



Acquired genetic modifications lead to clonal expansion over a background of polyclonal hematopoiesis



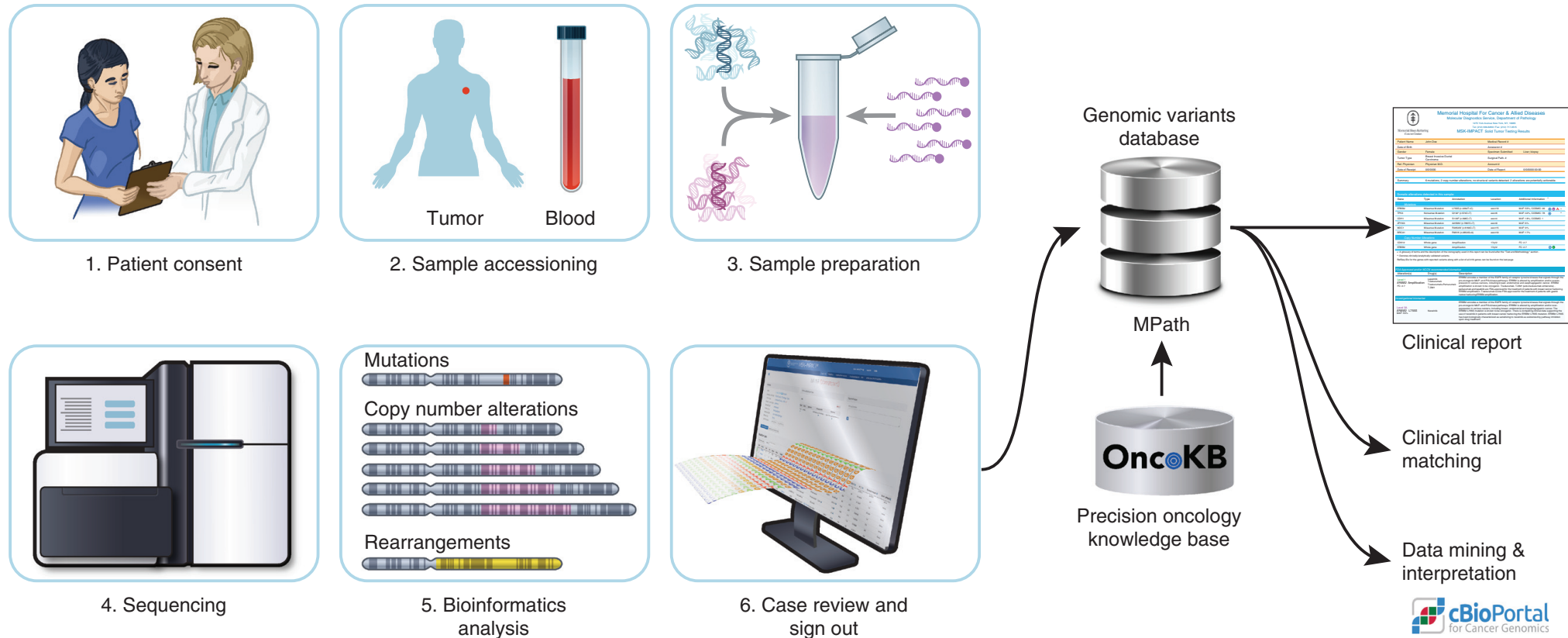
Main types of genetic changes are i) point mutations and small indels ii) mosaic chromosomal alterations, including mLOX and mLOY



Prevalence estimates of clonal expansions through different genetic alterations across an individual's lifespan (highly dependent on limit of detection of the assays)

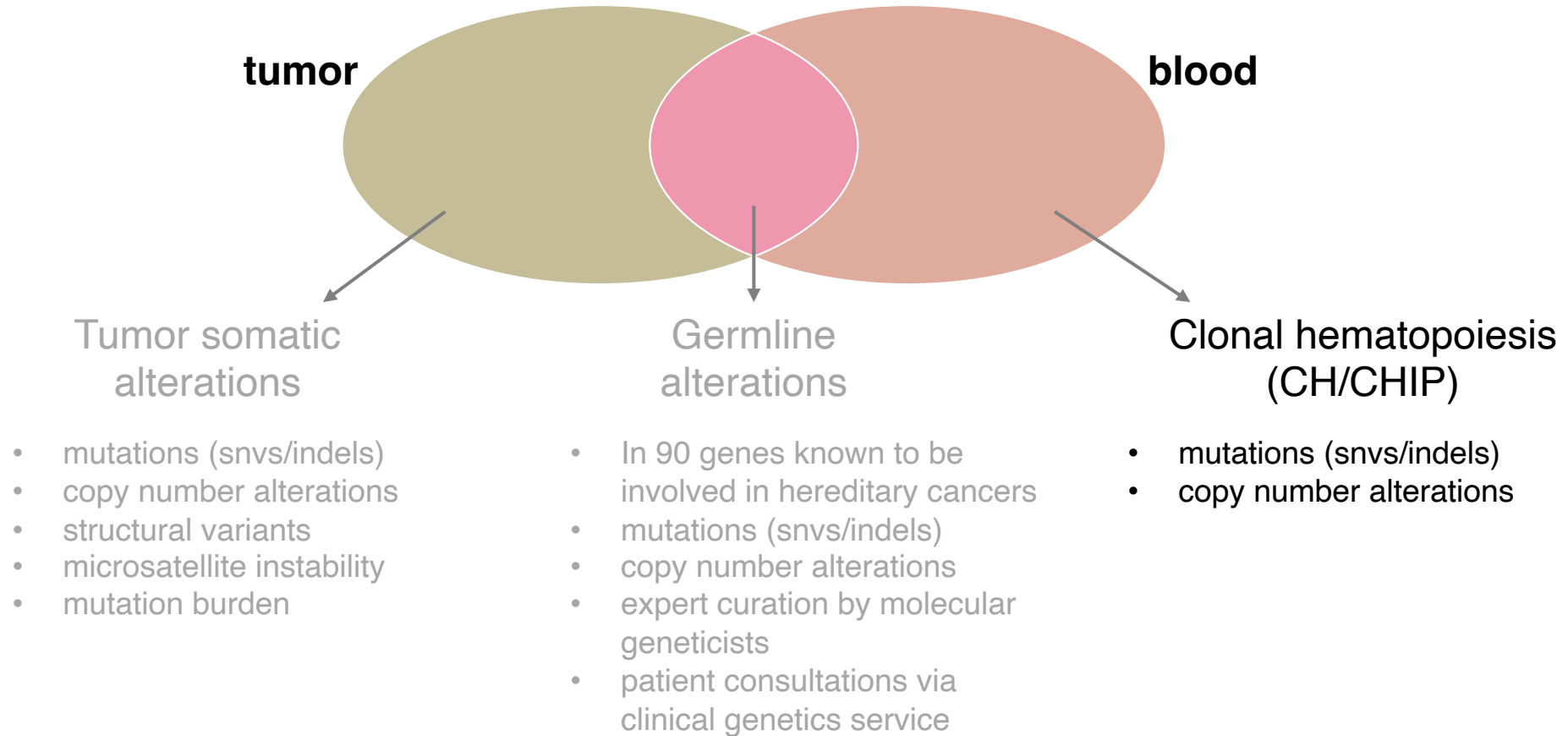
# Identification of CH mutations in solid tumor patients using MSK-IMPACT

Identifies mutations, copy number alterations, structural variants in **505 genes** as well as biomarkers that could predict treatment response such as **microsatellite instability (MSI) status** or **tumor mutation burden (TMB)**



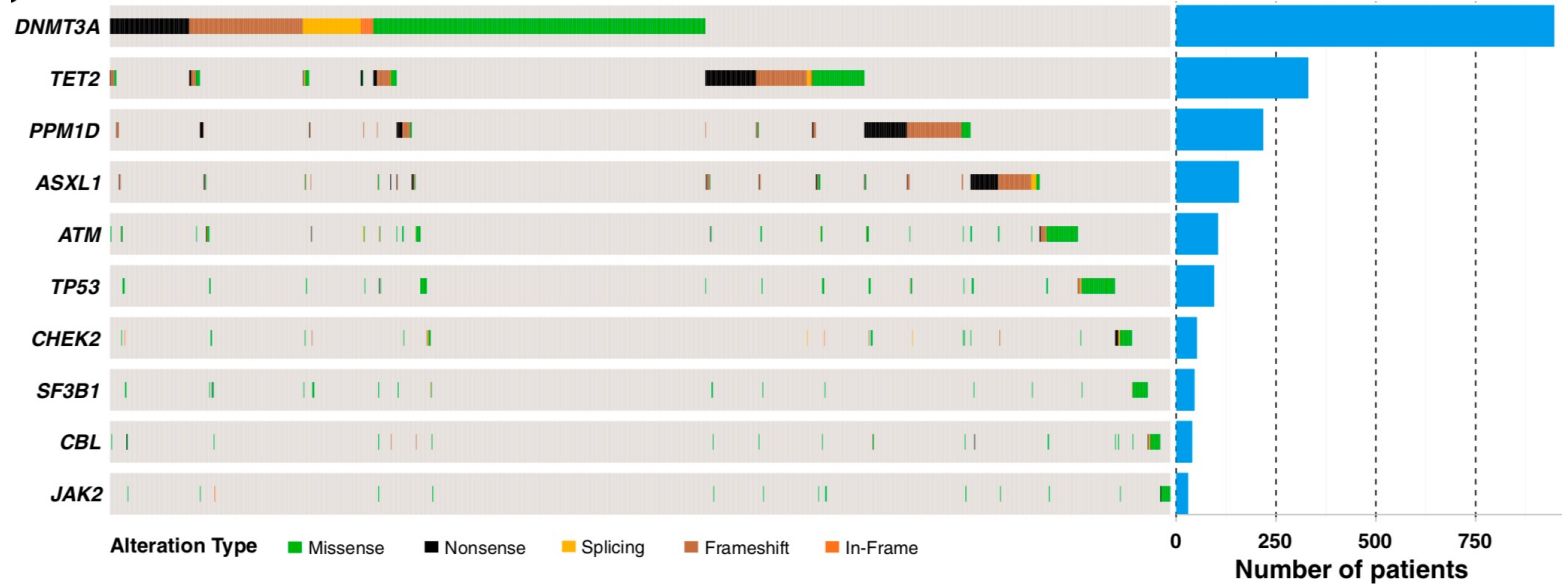
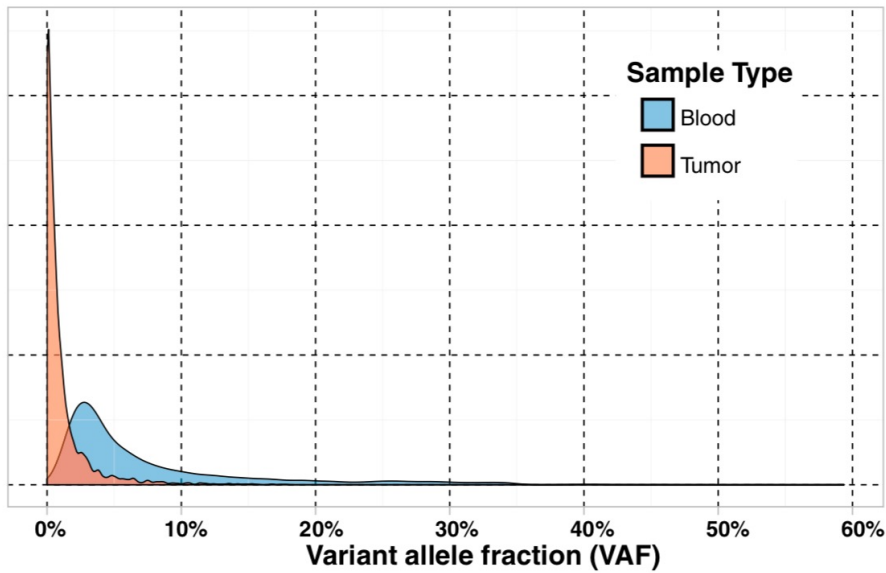
# Paired tumor & matched blood sequencing

Subtracting tumor mutations from the blood allows for identifications somatic mutations in the blood



# Clonal hematopoiesis is common in cancer patients

CH is observed in 1/4 patients, occurring in genes commonly mutated in heme malignancies

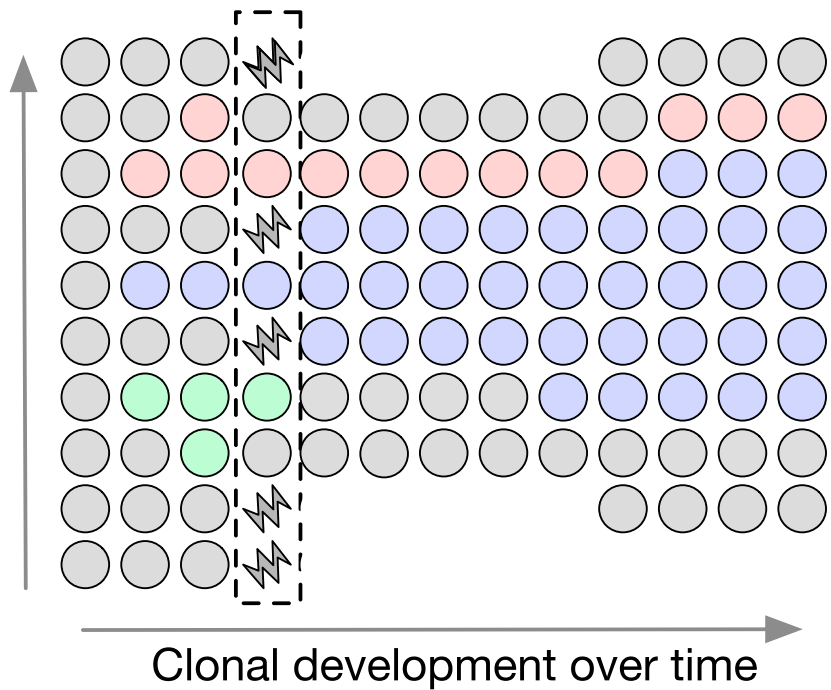


identified CH in ~ 24% of advanced cancer patients (n = 8,810)

# Clonal hematopoiesis

Oncologic therapy can shape further establishment and development of certain clones

Therapy-associated selective pressure



Bowman LB., *et al*, 2018 Cell Stem Cell Review

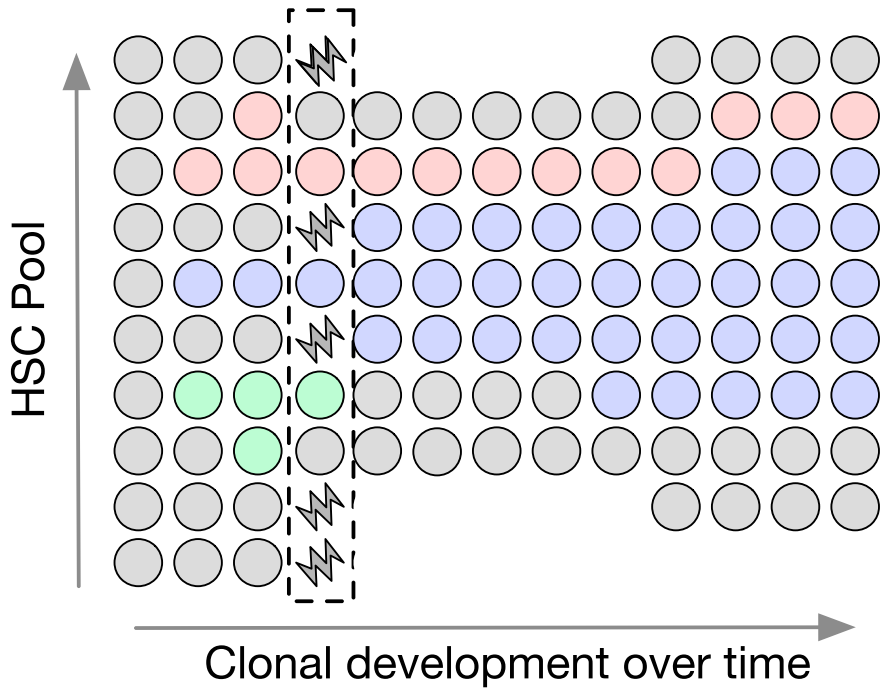
	Overall	CH	No CH	p value
Number of patients	5,649	1,353	4,296	–
Age (y)	58.3	66.1	55.8	<0.001
Gender (% male)	49	50	49	0.40
Associations with prior toxic exposures				
Current or former smoker (%)	46	53	44	<0.001
Prior chemotherapy (%)	64	64	64	0.89
Prior RT (%)	37	41	35	<0.001

Coombs CC, Zehir A., *et al*, 2017 Cell Stem Cell

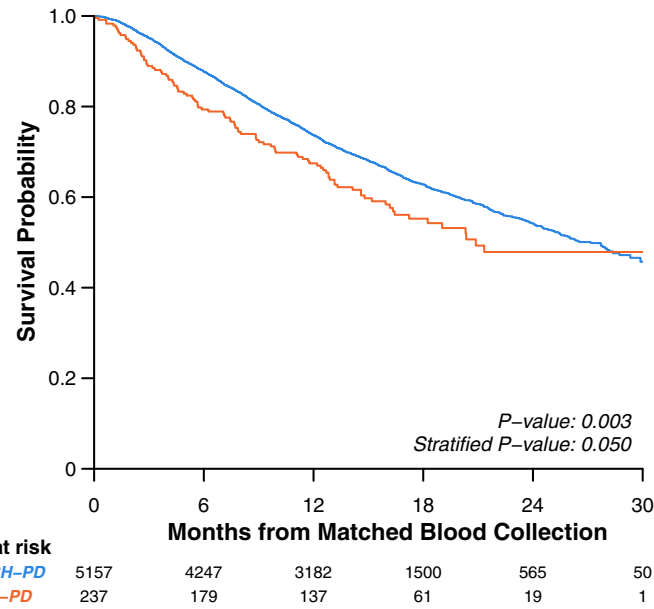
# Clonal hematopoiesis

Oncologic therapy can shape further establishment and development of certain clones

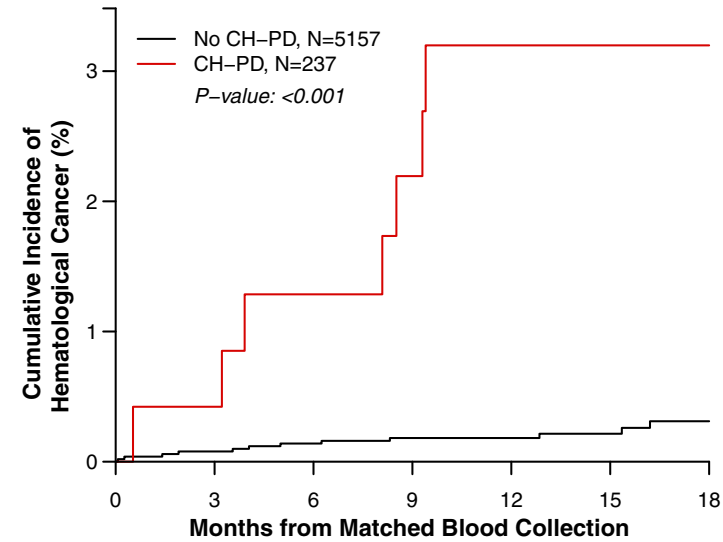
Therapy-associated selective pressure



Bowman LB., *et al*, 2018 Cell Stem Cell Review



Coombs CC, Zehir A., *et al*, 2017 Cell Stem Cell



# First CH clinic + real time CH reporting

These findings have led to the formation of the first CH clinic in the world, and we started reporting CH clinically

Somatic mutations detected in the blood (for investigational use - see assessment section):

Gene	Type	Alteration	Location	Additional Information
TET2	Missense	C1378Y ( <i>c.4133G&gt;A</i> )	exon 9	MAF: 13.7%
TET2	Frameshift Deletion	P333Hfs*14 ( <i>c.998del</i> )	exon 3	MAF: 19.7%
ASXL1	Frameshift Deletion	L731Yfs*13 ( <i>c.2191del</i> )	exon 12	MAF: 18.3%
PMS1	Frameshift Deletion	R760Sfs*19 ( <i>c.2280del</i> )	exon 10	MAF: 15.2%

MSK-IMPACT report

## MSK Opens New Clinic to Monitor People with a Genetic Risk for Developing Blood Cancer

By Julie Grisham, Tuesday, January 23, 2018



Ross Levine



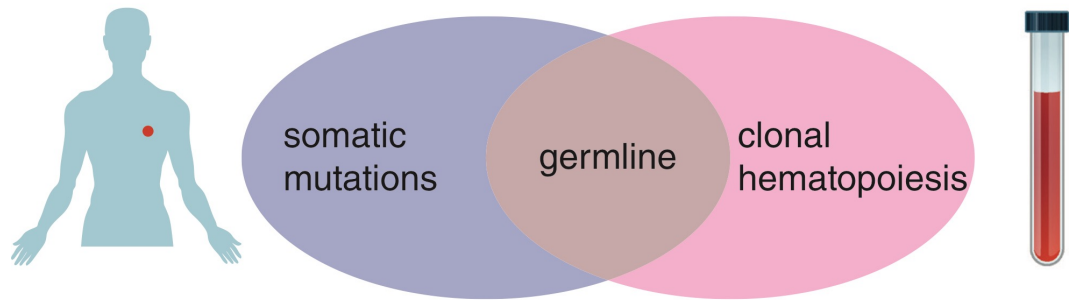
Kelly Bolton

CH clinic utilizes MSK-IMPACT data to identify patients who harbor CH mutations in real time. Patients are consulted and followed up if necessary



# Refining how oncologic therapy shapes CH in cancer patients

Identified CH in 24,354 patients and combined with deep phenotypic data on treatment



raw variant call set

- remove if a variant is present more than 3 times with  $\geq 1.5\%$  VAF in PON\*
- remove if a variant is present at all with  $\geq 2\%$  VAF in PON
- 1 bp deletions within a homopolymer stretch ( $\geq 3$ bp)
- SNVs completing a  $\geq 5$ bp homopolymer
- ins/dels in highly repetitive regions
- variants with strand bias (fisher test)

filter sequencing artifacts  
filter low quality samples  
require variants called by both VarDict and MuTect  
require  $VAF_{normal} > VAF_{tumor} * 1.5$  (if biopsy is from lymph node)  
 $VAF_{normal} > VAF_{tumor} * 2$  (otherwise)  
 $VAF \geq 2\% \mid AD \geq 10 \mid DP \geq 50$   
remove any variant if present in gnomad  $> 0.005$  MAF

final variant call set

\* PON: Pool of normals. 300 blood samples from persons  $< 20$  yrs old



Kelly Bolton

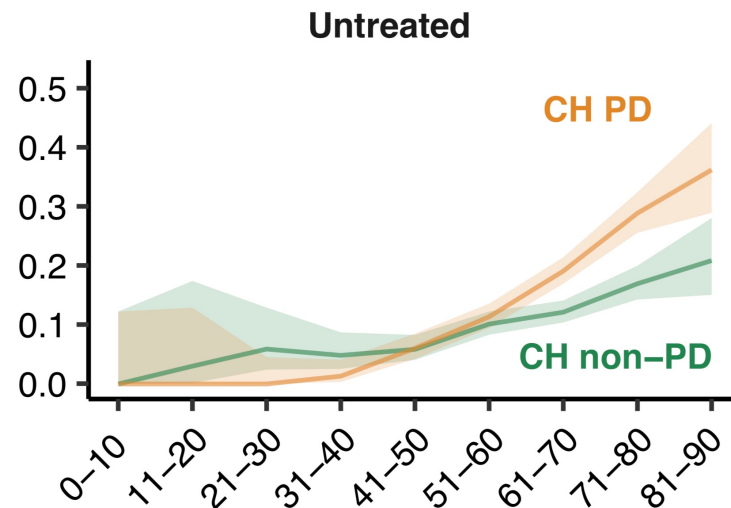
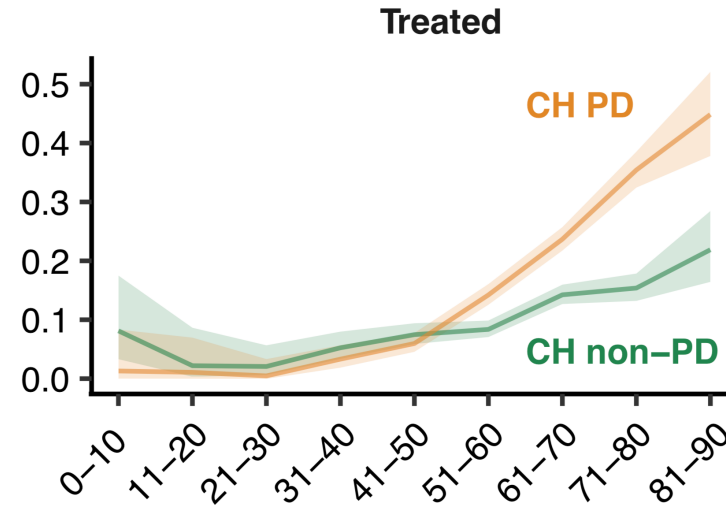
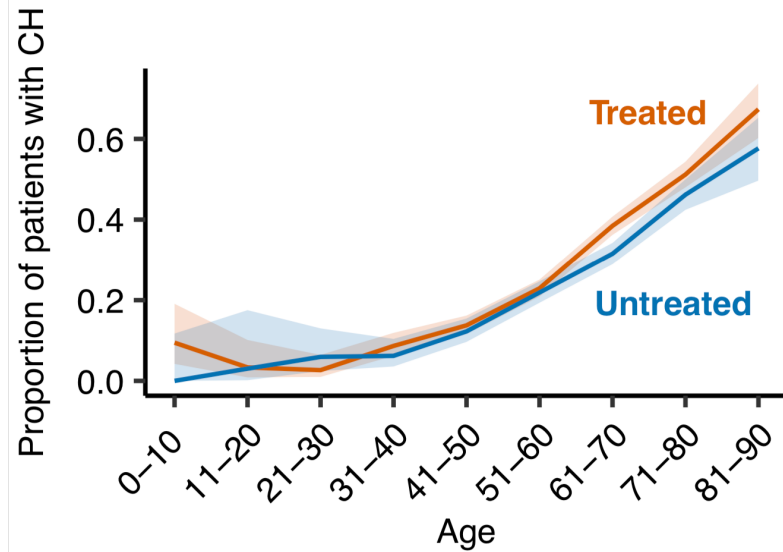
Ryan Ptashkin

Teng Gao

Elli Papaemmanuil

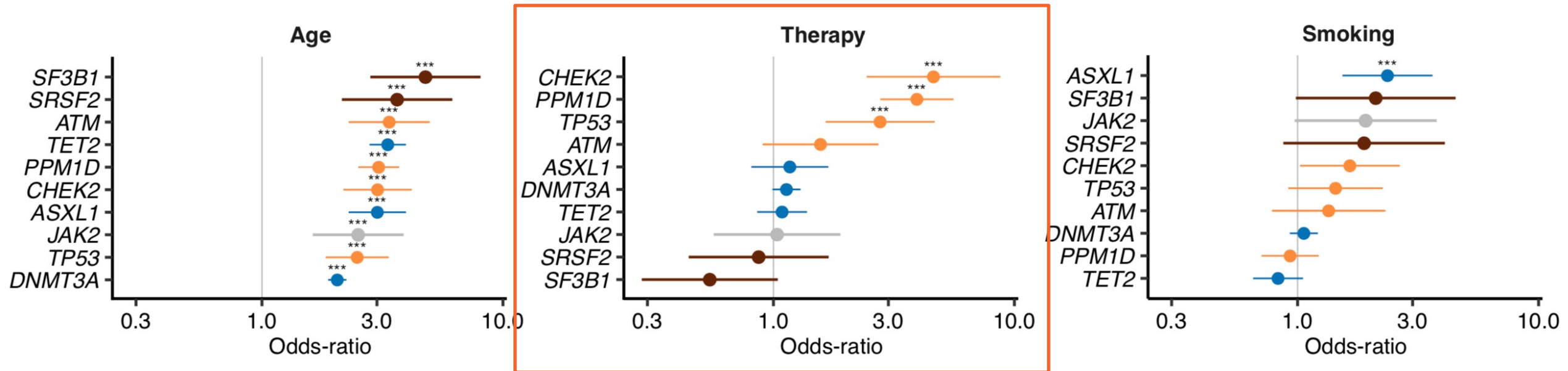
# Oncologic treatment affect CH rates across ages

CH in cancer driver genes (PD) shows the strongest selection relative to non-presumptive driver mutations

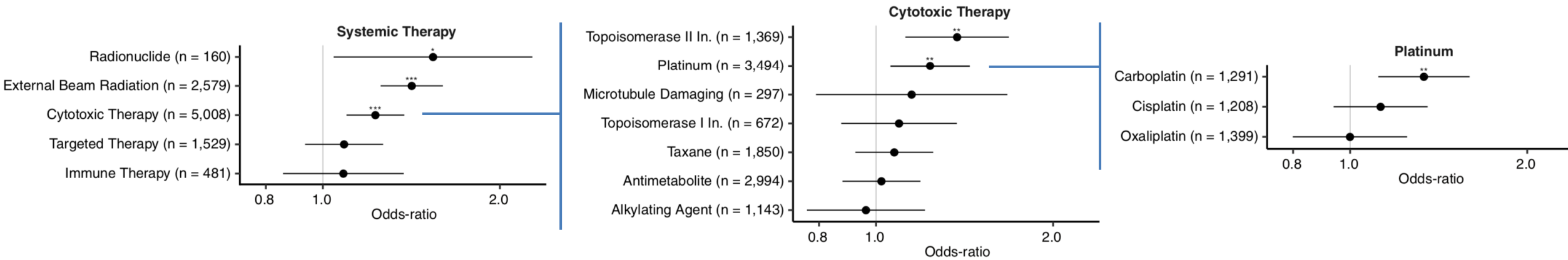


# Individual genes associate differently with selective pressures

While aging is associated with CH across all genes, therapy is specifically associated with CHEK2, PPM1D and TP53 and smoking is associated with ASXL1

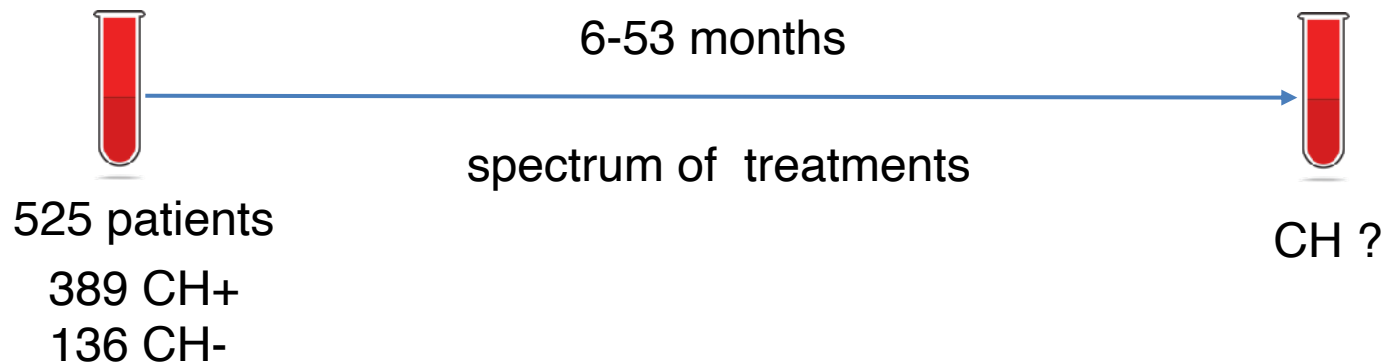


# CH-PD is associated with specific therapies

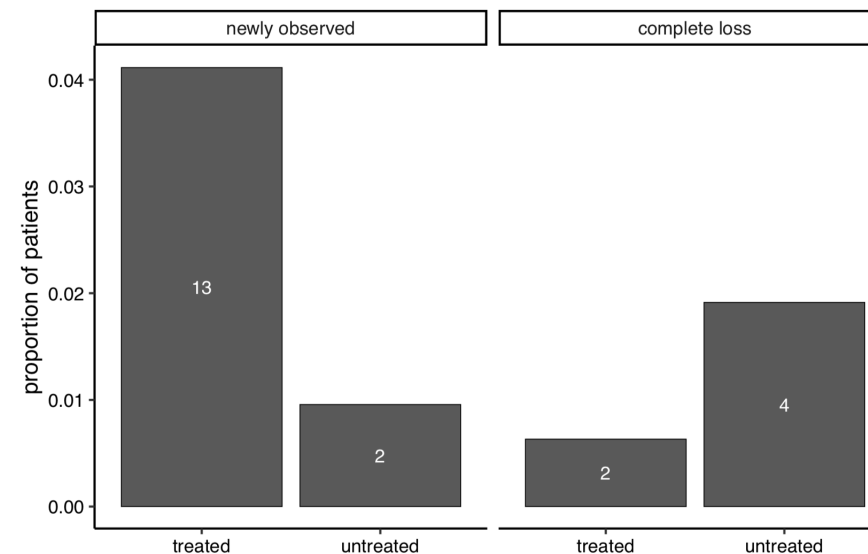


Multivariable logistic regression adjusted for age, gender, time from diagnosis to blood sequencing, race, all therapy subclasses

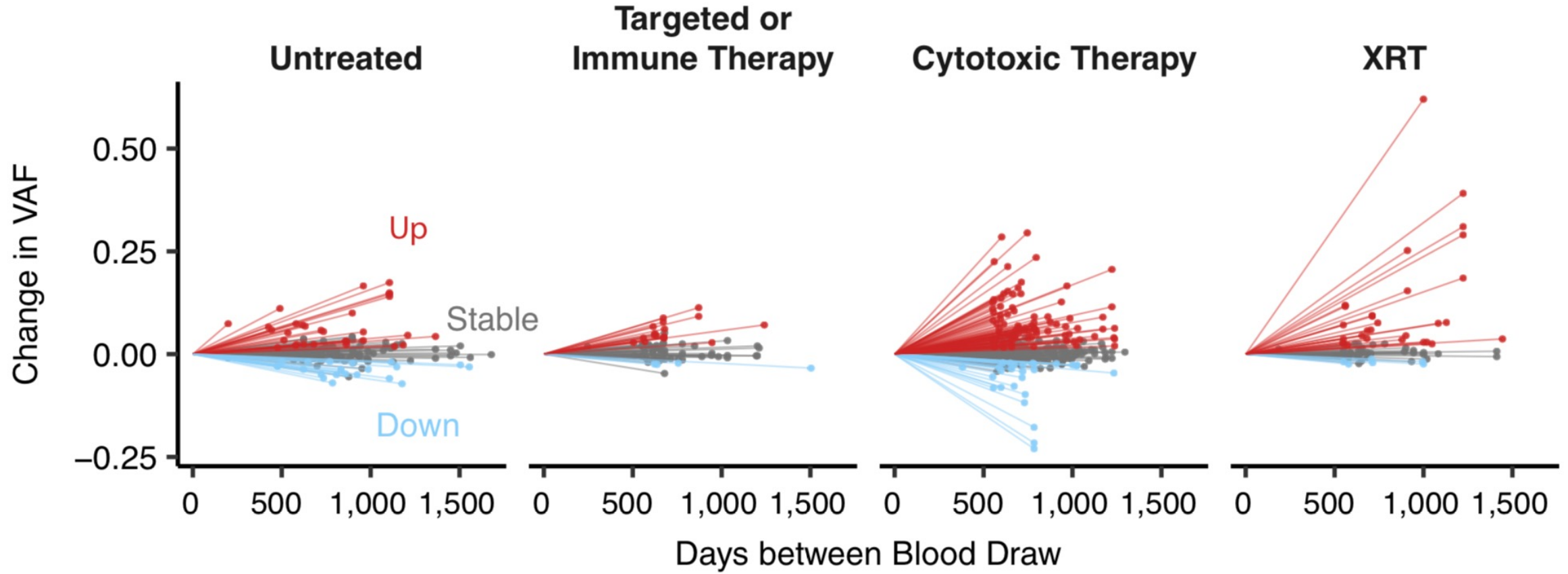
# Does ongoing treatment promote pre-existing CH or induce new mutations?



95% of the CH variants are detected at both time points

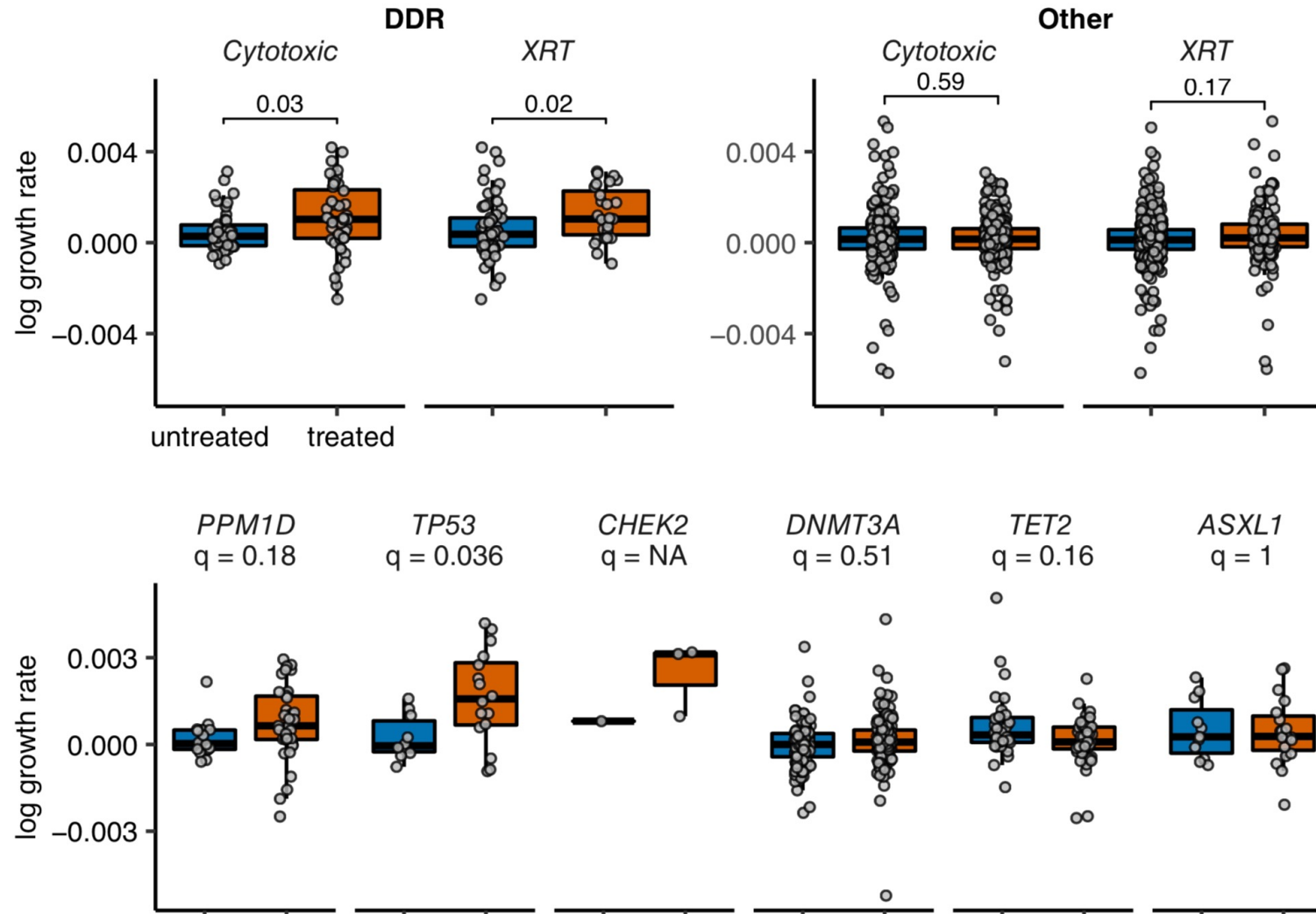


# Cytotoxic therapy and radiation promote growth of existing CH clones



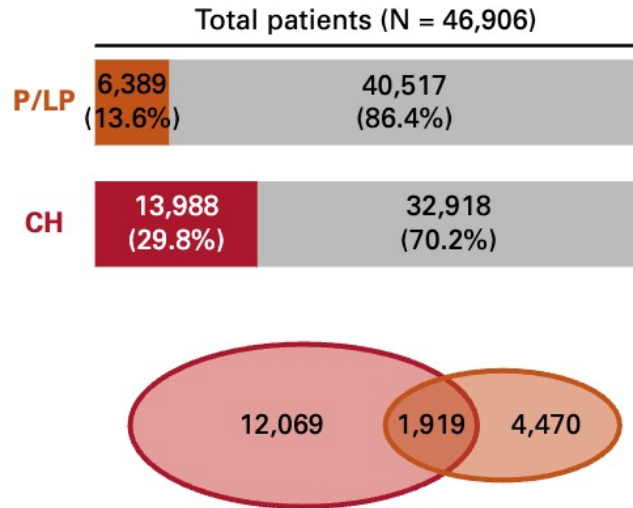
# Cytotoxic therapy and radiation promote growth of existing CH clones

*DDR pathway genes are specifically promoted*

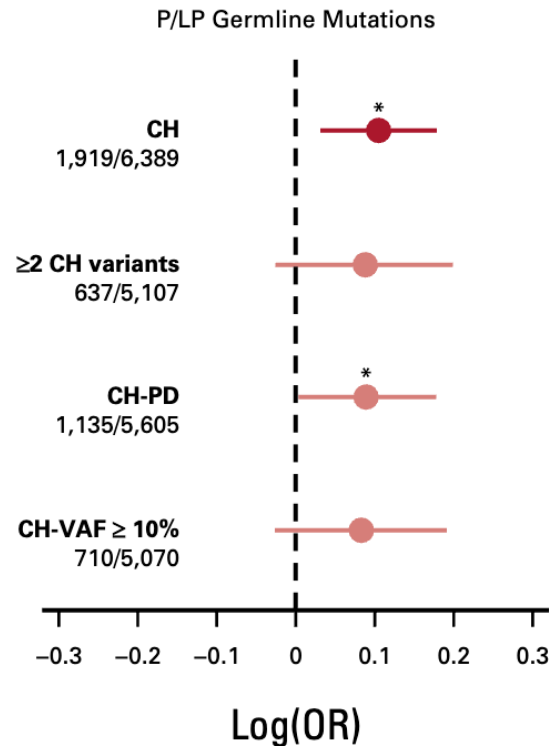


# CH associations with inherited cancer predisposition mutations

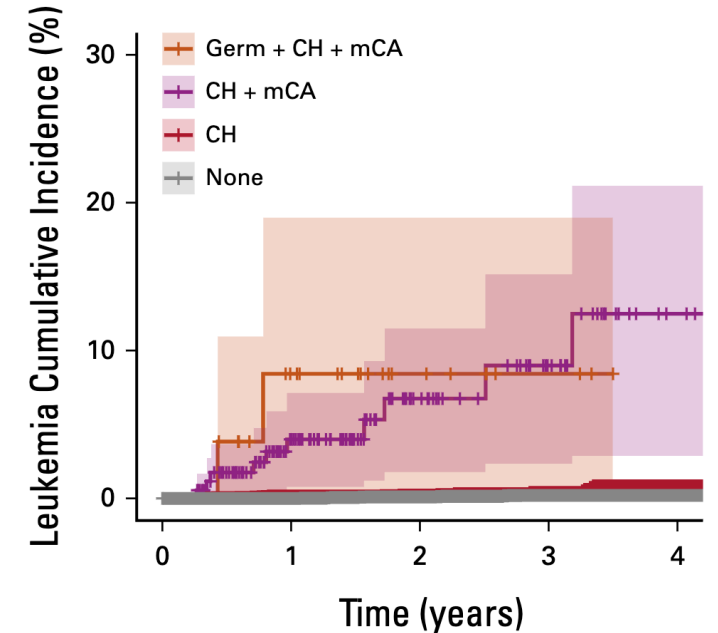
Cohort of 46,906 patients with CH and pathogenic/likely pathogenic germline mutations suggest additional population of patients who might need screening and monitoring



Amongst patients with P/LP and CH mutations, 10% of patients have both



Inherited cancer predisposition mutations are associated with presence of CH and CH-PD



Slightly increased leukemia incidence for Germline + CH + mCA – unfortunately small numbers



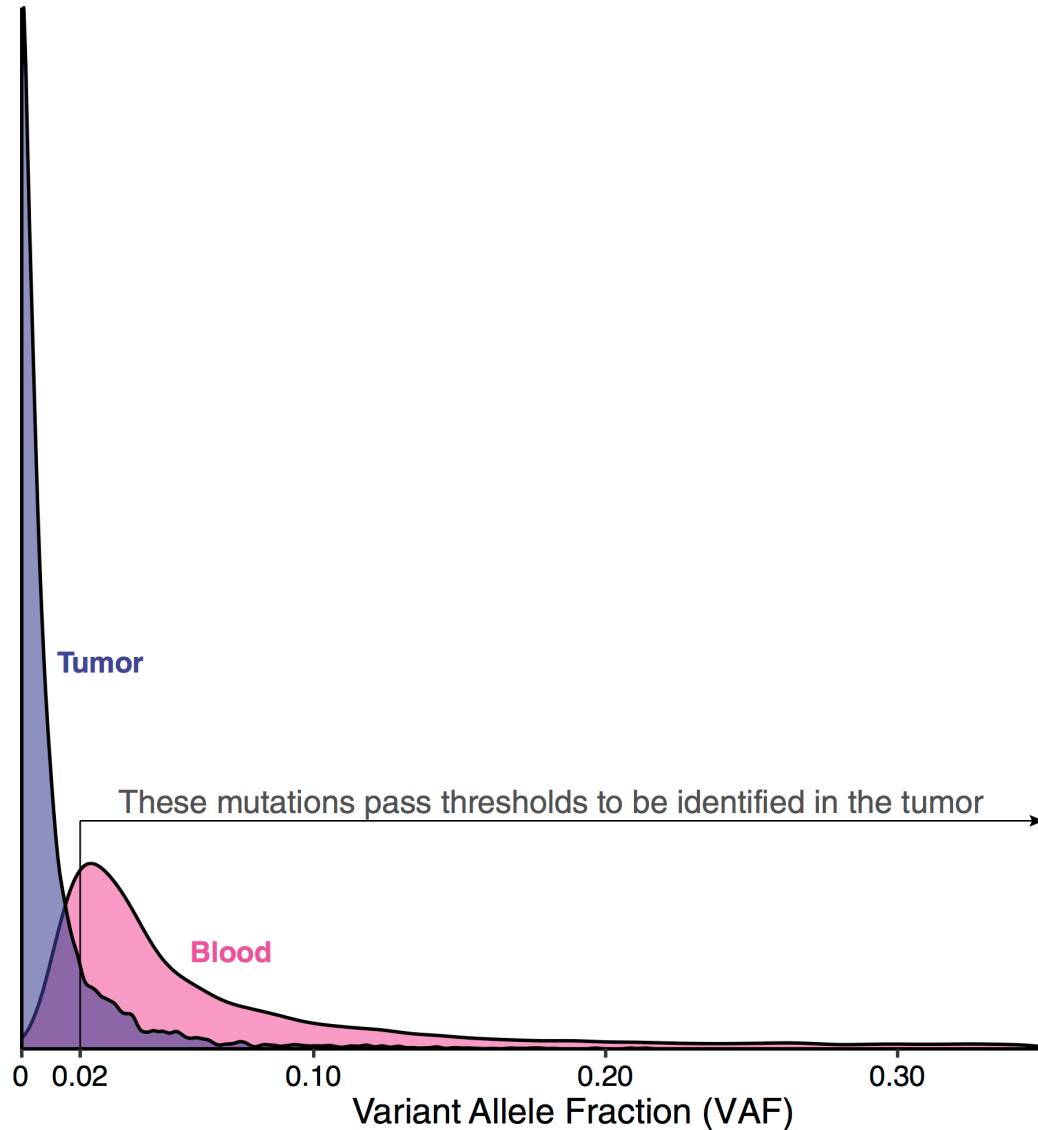


What are the implications of CH mutations in solid tumor diagnostics?

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# Identification of CH related mutations in the solid tumors

26.5% of patients have CH in their blood



Total # of patients = 17,469

Identified 7,608 CH mutations in 4,628 patients (26.5%)

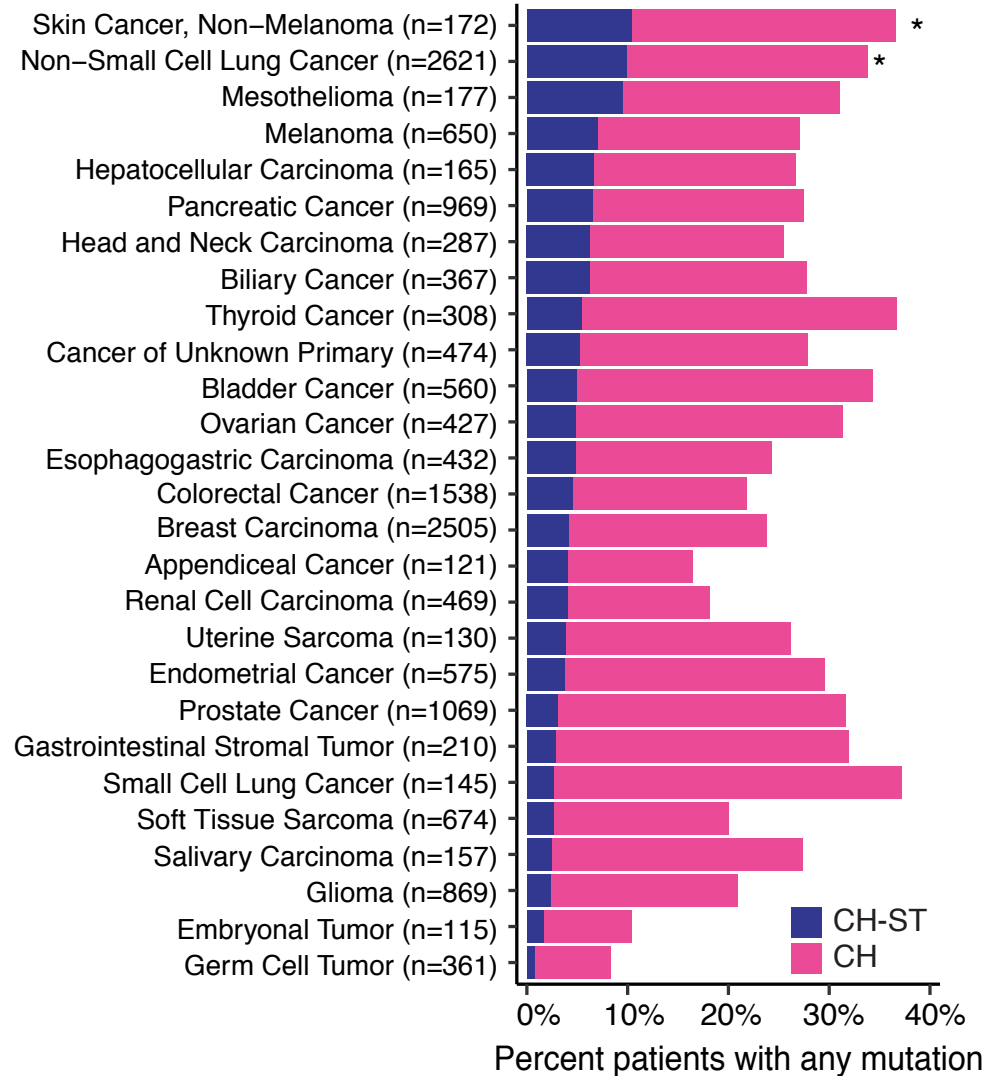
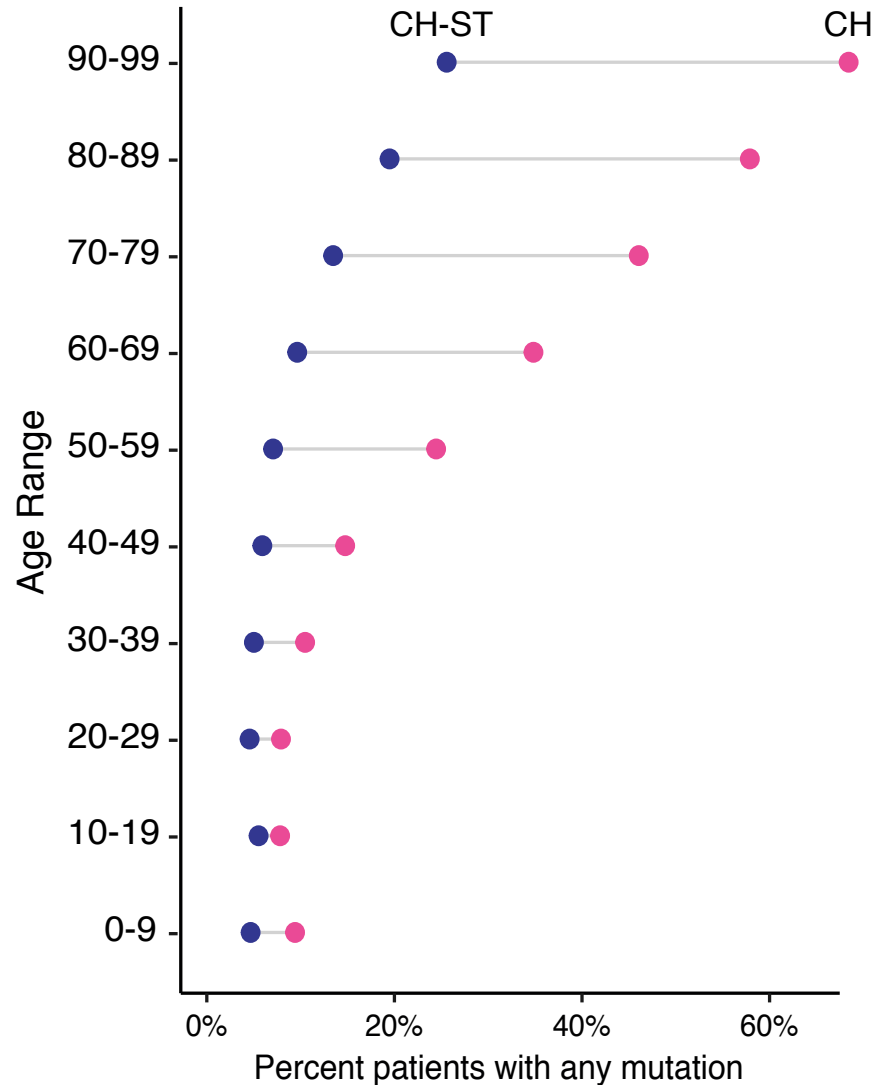
Identified 1,075 CH mutations in tumors of 912 patients (5.2%)



Ryan Ptashkin

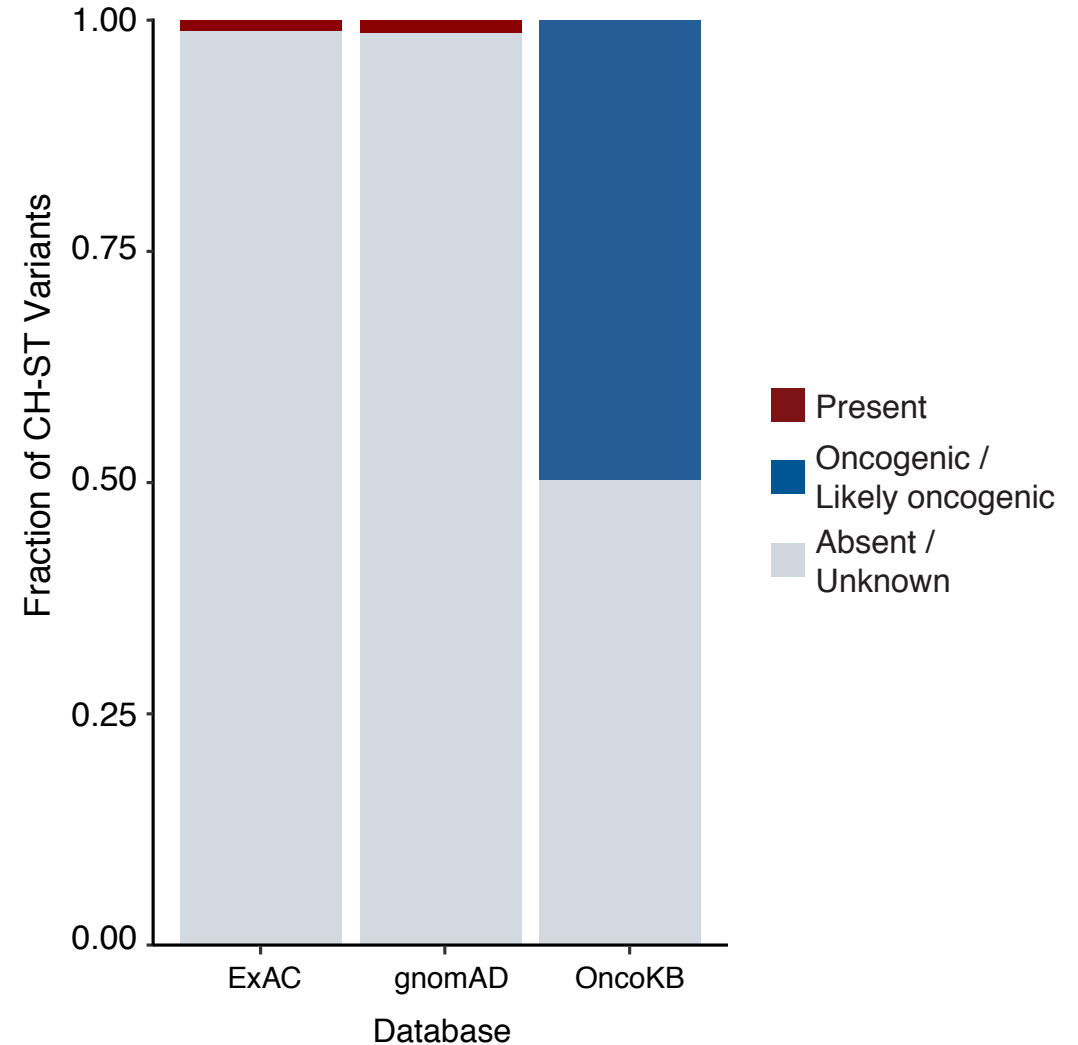
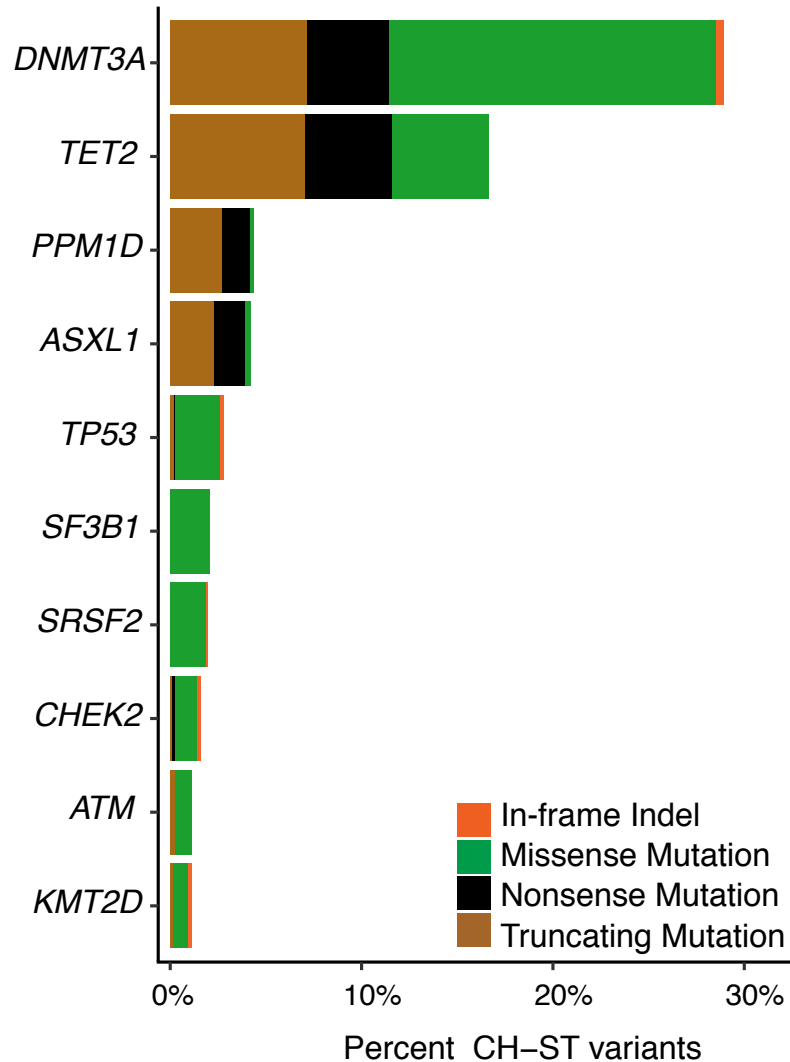
# CH mutations in solid tumors (CH-ST) is common

CH-ST incidence increase with age and varies by cancer type



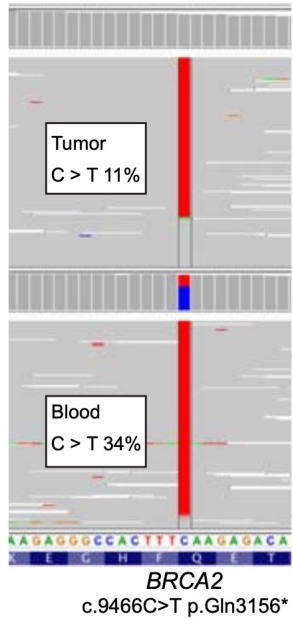
# CH-ST mutations are absent in population databases

CH-ST are observed in 206 genes, mostly absent in population databases and half are oncogenic/likely oncogenic



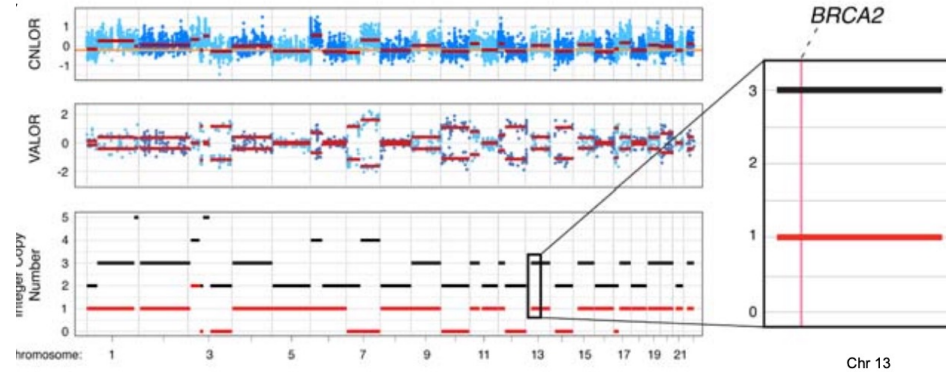
# CHIP as a confounding factor for germline mutations

BRCA2 pathogenic variant with implications for PARPi therapy

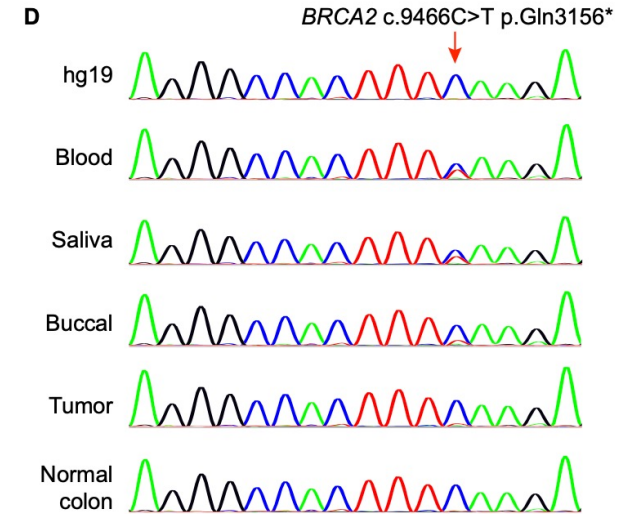


Looks somatic in tumor-only sequencing

Looks germline in blood-only sequencing



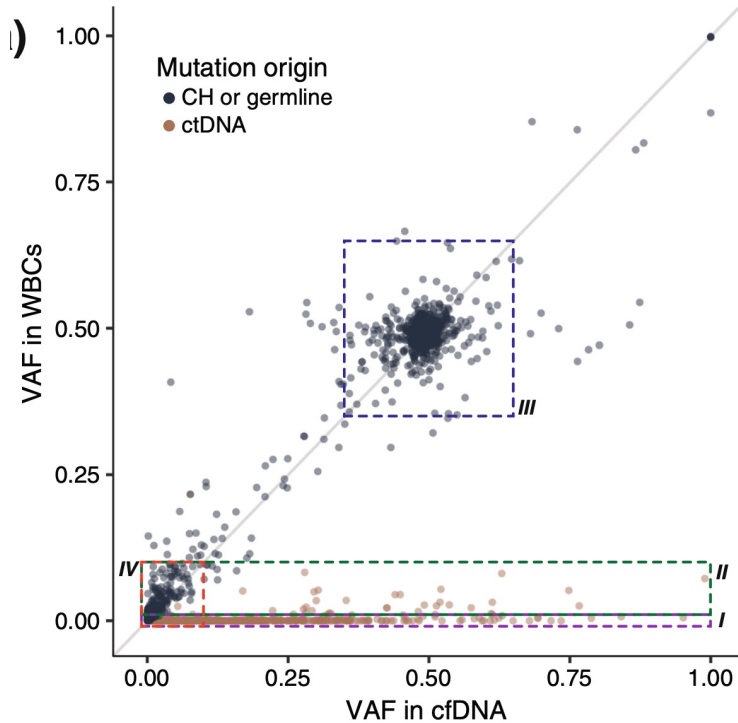
Tumor-blood data together suggests loss-of-heterozygosity in BRCA2 locus in tumor, however, both copies are present



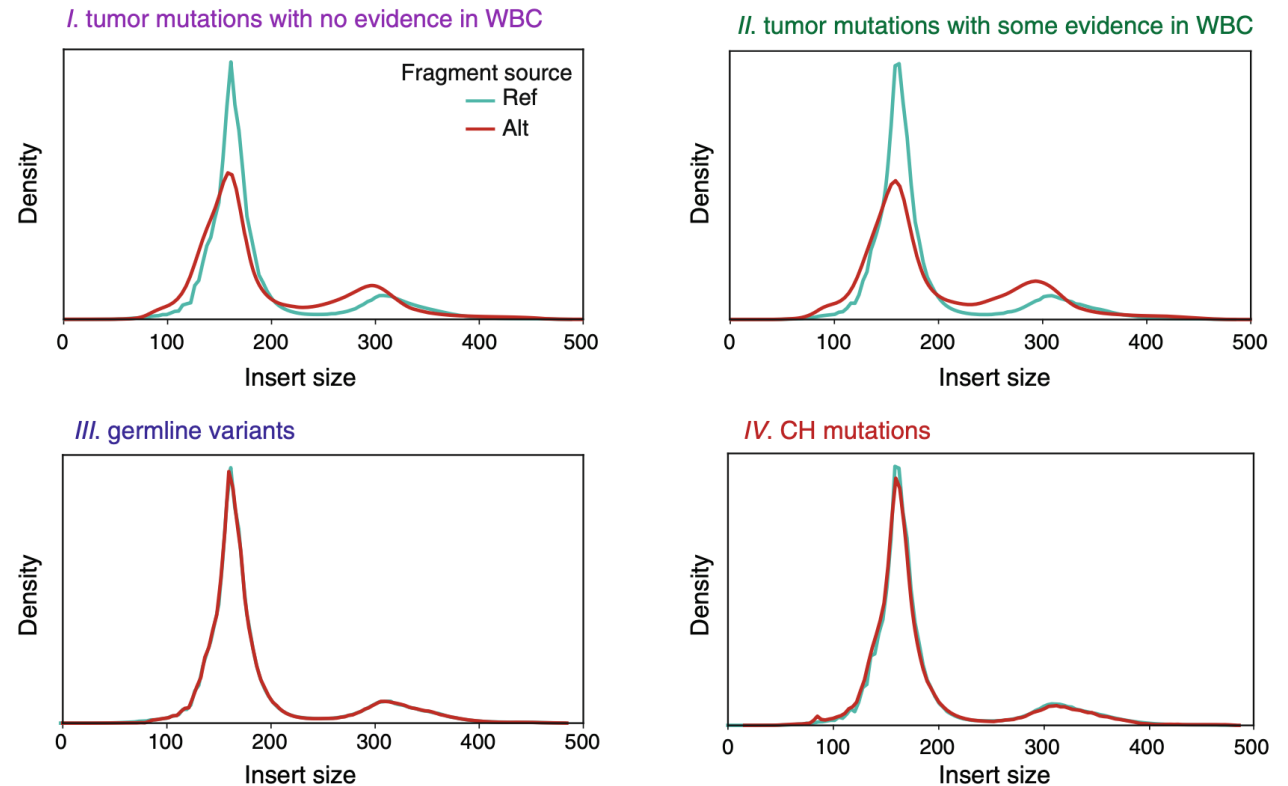
Sanger sequencing of different tissues show the mutation is a CH mutation

# Presence of CH mutations in liquid biopsies

Identification of genomic alterations in cell-free tumor-derived DNA (ctDNA) by sequencing white blood cells



Analysis of 617 plasma & WBC pairs, using MSK-ACCESS, shows up to 77% of variants removed through use of WBC data (< 10% VAF)

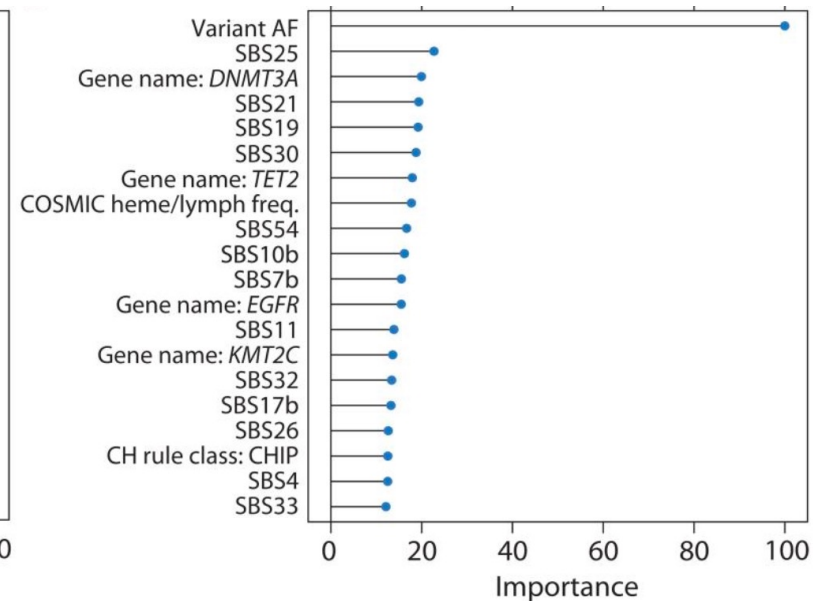
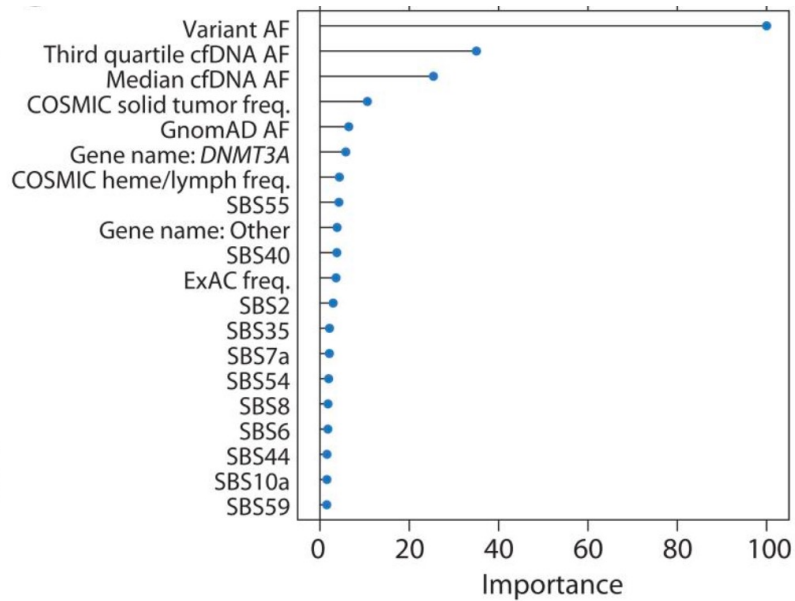
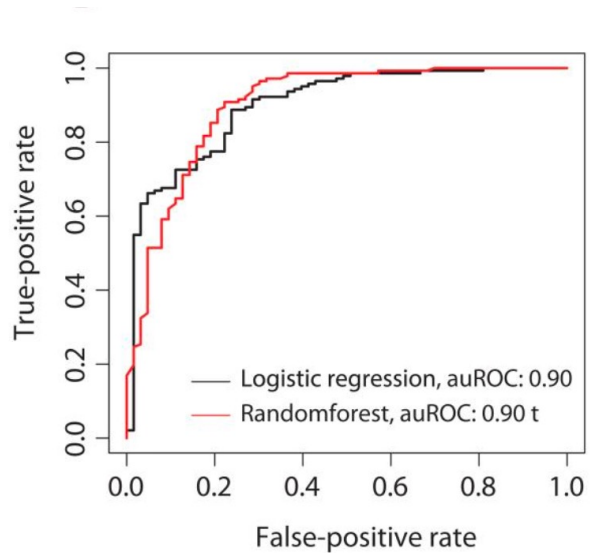


Fragments supporting tumor-derived mutations in plasma have shorter size distribution compared to the size of fragments supporting wild-type mutations.

Size distribution of fragments supporting germline and CH mutations do not differ from the background

# Presence of CH mutations in liquid biopsies

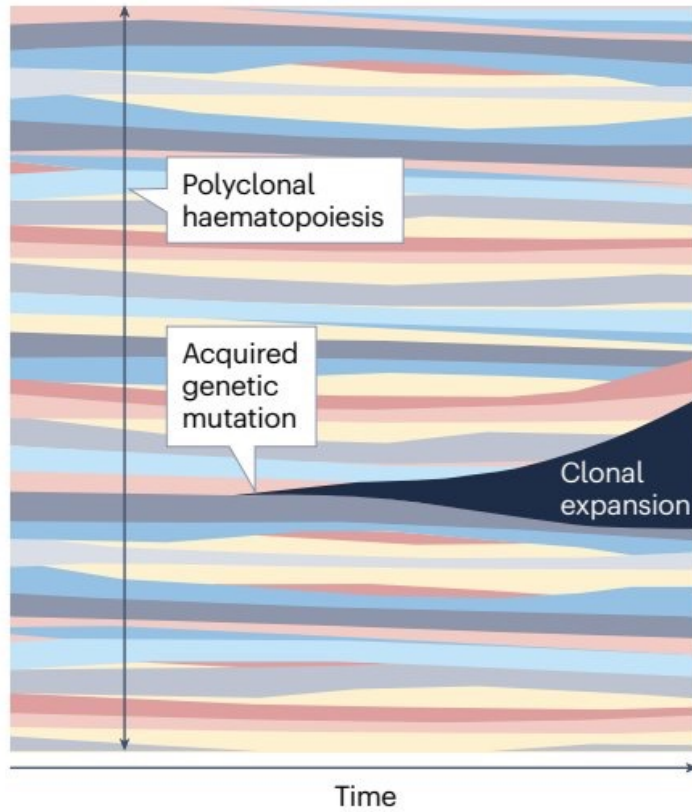
Machine learning approach to differentiate CH derived mutations from tumor mutations



Variables for 1,400 SNVs with known CHIP/somatic status are used to train a logistic regression or random forest classifier. Feature importance suggest mutation VAF, gene name and signature are critical. Data lacks use of fragment size information

# Conclusions

CH mutations can be confounders in diagnostic assays without appropriate controls and have implications for solid tumor patients



- i. noise for tumor-only sequencing (both ffpe & ctDNA)
- ii. noise for blood-only germline sequencing
- iii. signal for selecting patients at high-risk for hematological malignancies



# Remaining questions for the community

- 1) Can we use CH as a biomarker to identify high-risk cancer patients who might develop secondary malignancies? How would this look in practice? Can we treat these patients prophylactically?
- 2) What is the most optimal way to identify and remove CHIP derived noise in liquid biopsies in the absence of white blood cell sequencing?
- 3) Does presence of CH mutations lead to epigenetic changes thus confound methylation based liquid biopsy assays?
- 4) What is the extend of CHIP confounding in the setting of MRD?